

TM 1312 - VAGINALIS AGAR BASE

INTENDED USE

For isolation and differentiation of *Gardenerella vaginalis* from clinical samples.

PRODUCT SUMMARY AND EXPLANATION

Gardnerella vaginalis is a facultatively anaerobic gram-variable rod. It has been demonstrated to cause a wide variety of infections; however, it is most commonly recognized for its role as one of the organisms responsible for bacterial vaginosis (BV). BV is the most common cause of vaginitis and the most common infection encountered in the outpatient gynaecological setting. Originally Ellner et al developed a blood agar namely Columbia Agar for rapid growth of the haemolytic organisms with improved pigmentation and defined haemolytic reactions. Greenwood et al further modified this medium by increasing the peptone concentration and used human blood instead of sheep blood for the isolation and differentiation of *G. vaginalis* based on beta haemolysis. Vaginalis Agar Base is used for the isolation of *G. vaginalis* from vaginal discharges.

Typical colonies of *G. vaginalis* appear small and white coloured. This medium is recommended for determination of haemolytic reaction of *G. vaginalis* and not for other microorganisms. If the specimen is suspected to contain streptococci or other haemolytic microorganisms, then a Soyabean Casein Digest Agar (with 5% v/v sheep blood) plate should be inoculated parallel to this medium to ensure the haemolytic reaction.

COMPOSITION

Ingredients	Gms / Ltr
Casein enzymic hydrolysate	12.000
Peptic digest of animal tissue	15.000
Beef extract	3.000
Yeast extract	3.000
Corn starch	1.000
Sodium chloride	5.000
Agar	13.500

PRINCIPLE

Peptic digest of animal tissue, casein enzymic hydrolysate, yeast extract and beef extract provide nitrogenous compounds, carbon, sulphur, vitamin B complex and trace ingredients required for growth. Cornstarch serves as the energy source. Blood supplies additional nutrients and also aids in identification.

INSTRUCTION FOR USE

- Dissolve 52.5 grams in 950 ml distilled water.
- Heat to boiling to dissolve the medium completely.
- Dispense in 95 ml amounts and sterilize by autoclaving at 15 psi pressure (121°C) for 15 minutes.
- Cool to around 50-55°C and aseptically add 5 ml of sterile anticoagulated human blood to every 95 ml sterile basal medium.
- Mix well and pour into sterile Petri plates.

QUALITY CONTROL SPECIFICATIONS



Appearance of Powder : Cream to yellow homogeneous free flowing powder.
Appearance of prepared medium : Basal medium: Yellow coloured clear to slightly opalescent gel. After addition of 5% v/v sterile anticoagulated human blood, cherry red coloured, opaque gel forms in Petri plates.
pH (at 25°C) : 7.4±0.2

INTERPRETATION

Cultural characteristics observed in an aerobic atmosphere containing 3-10% CO₂ with added 5% v/v sterile anticoagulated human blood after an incubation.

Microorganism	ATCC	Inoculum (CFU/ml)	Growth	Recovery	Haemolysis	Incubation Temperature	Incubation Period
<i>Gardnerella vaginalis</i>	14018	50-100	Good-luxuriant	>=50%	Beta (diffused)	35-37°C	48 Hours

PACKAGING:

In pack size of 500 gm bottles.

STORAGE

Dehydrated powder, hygroscopic in nature, store in a dry place, in tightly-sealed containers between 25-30°C and protect from direct sunlight. Under optimal conditions, the medium has a shelf life of 4 years. When the container is opened for the first time, note the time and date on the label space provided on the container. After the desired amount of medium has been taken out replace the cap tightly to protect from hydration.

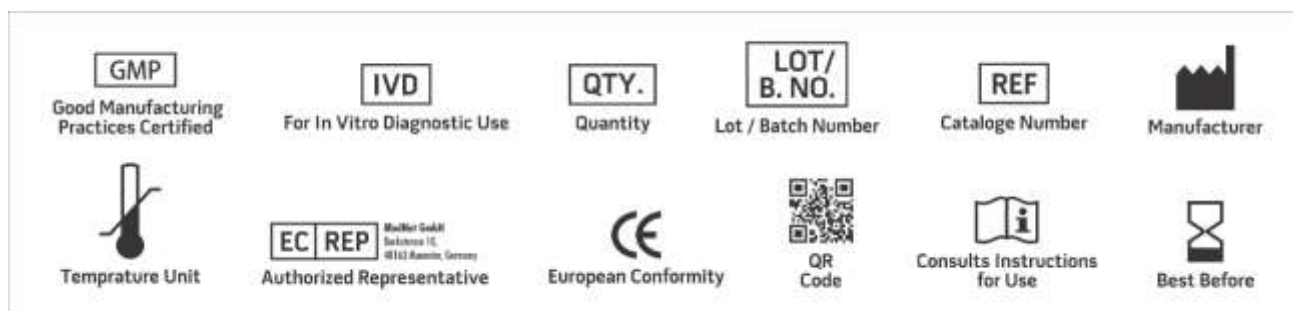
Product Deterioration: Do not use if they show evidence of microbial contamination, discoloration, drying or any other signs of deterioration.

DISPOSAL

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

REFERENCES

1. Ellner P. D., Stoessel C. J., Drakeford E., Vasi F., 1966, Am. J. Clin. Pathol., 45: 502.
2. Greenwood J. R., Martin M. J., Mack E. G., 1977, Health Lab. Sci., 14: 102.
3. Greenwood J. R. and Pickett M. J., 1980, Int. J. Syst. Bacteriol., 30: 170.
4. Piot P., Van Dyck E., Goodfellow M., Falkow S., 1980, J. Gen. Microbiol., 119: 373.
5. MacFaddin J. F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. 1, Williams and Wilkins, Baltimore



NOTE: Please consult the Material Safety Data Sheet for information regarding hazards and safe handling Practices.



***For Lab Use Only**
Revision: 08 Nov., 2019

